Antiproteases in Preventing the Invasive Potential of Pancreatic Cancer Cells

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Summary

The process of tumor progression and metastasis involves degradation of the extracellular matrix and is governed by an intricate balance of proteases, their activators and their inhibitors, in which malignant cells are permitted to infiltrate the adjacent structures and gain access to lymph and blood vessels. These proteases can be broadly categorized into three families: matrix metalloproteinases, serine proteinases and cysteine proteinases, all of which have all been implicated in these processes. The presence of neural invasion is often considered to be a poor prognostic sign; however, the cellular mechanisms underlying this propensity for perineural invasion are unknown. We recently researched the relationship between the glial cell line-derived neurotrophic factor and perineural invasion by human pancreatic cancer cells. We also confirmed that NF-kappa B is a part of the signaling pathway from the glial cell linederived neurotrophic factor in pancreatic cancer cells, and documented the inhibitory effect of gabexate mesilate, a wellknown non-physiological synthetic serine protease inhibitor, for pancreatic cancer invasion. Recent studies on the role of proteases and protease inhibitors in pancreatic cancer invasion are also reviewed.

Introduction

Of all gastrointestinal carcinomas, pancreatic cancer has the most unfavorable prognosis, and many patients die from liver metastasis or local recurrence because pancreatic cancer frequently and rapidly invades the surrounding tissue, such as the lymph nodes and neural plexuses. In particular, the presence of neural invasion is widely considered to be a poor prognostic sign [1, 2, 3], but the cellular mechanisms underlying this propensity for perineural invasion remain unknown.

Recent studies report that transforminggrowth-factor-beta (TGF-beta) is implicated in the metastatic and invasive potential of pancreatic cancer [4]. The glial cell linederived neurotrophic factor (GDNF), a distantly related member of the TGF-beta family which was originally purified from the B49 glial cell line [5], is a potent survival factor for dopaminergic neurons motoneurons [5, 6]. We recently investigated relationship **GDNF** between perineural invasion by human pancreatic cancer cells. The results revealed that the invasive potential was increased by GDNF in human pancreatic cancer cell lines; we confirmed that nuclear factor kappaB (NFkappa B) is a part of the signaling pathway from GDNF in human pancreatic cancer cells [7, 8, 9]. These results indicate that the NF-

kappa B inhibitor has a marked influence on pancreatic cancer invasion and suggest that these drugs may have an application in the prevention and treatment of pancreatic cancer. Gabexate mesilate is a well-known nonphysiological synthetic serine protease inhibitor. Gabexate mesilate inhibits various serine proteases such as trypsin, plasmin and thrombin, and has been used in Japan for the of acute pancreatitis treatment and disseminated intravascular coagulation [10]. Gabexate mesilate is also commonly used endoscopic retrograde cholangiopancreatography and pancreatic cancer surgery to prevent post-procedure postoperative pancreatitis [11].

It has recently been reported that gabexate mesilate inhibits NF-kappa B activation in human monocytes and human umbilical vein endothelial cells (HUVECs) [12, 13]. The transcription factor NF-kappa B is a homodimer or heterodimer composed of members of the rel/NF-kappa B protein family, including Rel A, Rel B, c-Rel, p50/p105, and p52/p110. NF-kappa B has various functions in cancer cells, including the prevention of apoptosis [14] and the promotion of chemoresistance [15, 16], cell invasion and metastases [17, 18]. In several types of malignancies, NF-kappa B is activated constitutively [19, 20, 21]. The inhibition of NF-kappa B activity by one of its inhibitors, such as a proteasome inhibitor, sulfasalazine or salicylate, reduces NF-kappa B-dependent protection from apoptosis [15, 22, 23, 24].

While several reports describe the role of serine protease inhibitors in cancer, little is known about their influence on invasive potential in human pancreatic cancer cells. In the present study, we investigate the influence of serine protease inhibitors on NF-kappa B activation and invasive potential in human pancreatic cancer, and conduct a review of recent studies on the role of proteases and protease inhibitors in pancreatic cancer invasion.

Serine Protease Inhibitors and Our Findings

Serine protease inhibitors can be categorized into physiological inhibitors which are naturally present in tissues and nonphysiological inhibitors which are either produced by micro-organisms or chemically synthesized. Physiological serine protease inhibitors belong to the superfamily of serpins which includes antithrombin III, PAI-1 and PAI-2 [25], among others. Serine proteases are known to have various effects on cancer cells. One such serine protease is the urokinase-type plasminogen activator (uPA). Plasmin has the ability of activating the matrix metalloproteinases necessary basement membrane degradation [26]. uPA, which activates plasmin, is overexpressed in pancreatic cancer cells and has been shown to be involved in tumor invasion and metastasis [27]. The concomitant overexpression of uPA and uPA receptor (uPAR) was found to be associated with shorter survival in pancreatic cancer Sawai patients [28]. et demonstrated that IL-1alpha can induce selective upregulation of uPA/uPAR in pancreatic cancer cells and that inhibitory antibodies against uPAR can reduce the invasive potential of pancreatic cancer cells [29]. Sawai et al. reported that peroxisome proliferators-activated receptor (PPAR)gamma ligands, which are currently in clinical use as antidiabetic drugs, decrease pancreatic cancer invasion via modulation of the plasminogen activator system [30].

We previously reported that pancreatic cancer cell lines have both GDNF receptors (GFRalpha-1 and Ret), and that the invasive capacity of human pancreatic cancer cell lines is increased by GDNF or co-cultivation with human glioma cells T98G or A172 [7, 8]. We also reported that NF-kappa B is a part of the signaling pathway from GDNF in human pancreatic cancer cells and that NF-kappa B activity is strongly correlated with its invasive potential [9]. In agreement with our results, Zhang *et al.* reported that the epidermal

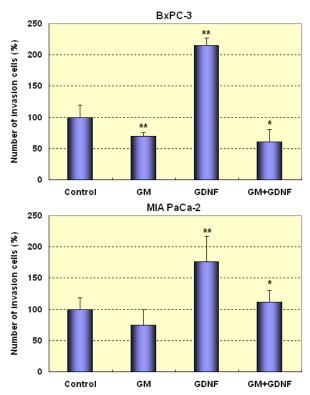


Figure 1. The effect of gabexate mesilate and GDNF on pancreatic cancer invasion. Using the Biocoat Matrigel Invasion Chamber system (BD Biosciences, Bedford, MA, USA), cells $(1\times10^5/\text{well})$ were incubated on the upper component with gabexate mesilate (GM: $100~\mu\text{M}$) and GDNF (100~ng/mL) for 24 hours. Statistical significance was analyzed by non-repeated ANOVA with an SNK test. Bars indicate SD, **P<0.01 vs. controls, and *P<0.01 vs. GDNF only.

growth factor (EGF) gives rise to matrix metalloproteinase-9 (MMP-9), uPA induction and the invasiveness of pancreatic cancer through the NF-kappa B pathway [31].

We recently investigated the influence of the synthetic serine protease inhibitor gabexate mesilate on GDNF-induced NF-kappa B activation and invasive potential in two human pancreatic cancer cells (BxPC-3, MIA) PaCa-2) using a dual-luciferase reporter assay and an in vitro invasion assay. The results demonstrated that gabexate mesilate prevents GDNF-induced NF-kappa B activity in a dose-dependent manner and significantly inhibits invasive ability in two pancreatic cancer cell lines (Figures 1, 2). It is unclear, however, how gabexate mesilate inhibits NFkappa B activation. In human monocytes and HUVECs, gabexate mesilate inhibits IkappaB phosphorylation and degradation as well as NF-kappa B localization to the nucleus [12, 13]. Gabexate mesilate may inhibit the degradation of I-kappaB in a manner similar to that of other proteasome inhibitors such as MG132 or PS-341 by inhibiting the 26S proteasome [15, 22, 32]. Uchima et al. reported that gabexate mesilate could inhibit pancreatic cancer invasion by directly antagonizing the activities of uPA and tumor-associated trypsinogen (TAT). The uPA-promoter contains an NF-kappa B binding site which directly mediates the induction of uPA expression [33]; therefore, we consider that gabexate mesilate could inhibit pancreatic cancer invasion not only by antagonizing the activities of uPA and TAT, but also by suppressing uPA production

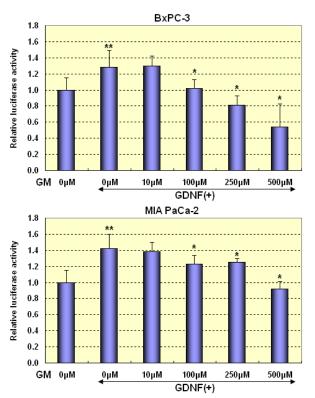


Figure 2. The effect of gabexate mesilate and GDNF on NF-kappa B activity. Cells $(5\times10^4/\text{well})$ were exposed to GDNF (100 ng/mL) and gabexate mesilate (GM: 0-500 μM) for 6 hours, after which relative luciferase activity was measured. GDNF significantly increased the relative luciferase activity of both cell lines, and the increased NF-kappa B activity was suppressed by pre-treatment with gabexate mesilate at concentrations higher than 100 μM. Statistical significance was analyzed by non-repeated ANOVA with Dunnett's test. Bars indicate SD; **P<0.01 vs. controls; *P<0.01 vs. GDNF only.

(Figure 3). Furthermore, we reported that gabexate mesilate could prevent NF-kappa B activation and increase TNF-alpha mediated apoptosis in human pancreatic cancer cells by suppressing the NF-kappa B signaling pathway [34]. These findings further support the notion that serine protease inhibitors are potentially promising therapeutic targets in pancreatic cancer.

Matrix Metalloproteinases and Their Inhibitors

Matrix metalloproteinases (MMPs) are a family of zinc-dependent proteolytic enzymes capable of degrading the extracellular matrix (ECM) which plays a critical role during cancer cell invasion [35, 36, 37]; 18 different subtypes have recently been identified [38]. Recent studies report that MMP-2 and/or MMP-9 were overexpressed in pancreatic cancer; this expression strongly correlated with the presence of the desmoplastic reaction and the lymphoid reaction [39, 40]. MMP-7 is also involved in cell dissociation and the subsequent invasion of pancreatic cancer [41]. A variety of stimuli (cytokines, growth factors, cellular stress) are reported to upregulate the expression of MMPs; for example, Okada et al. reported that GDNF upregulates the expression and activation of MMP-9 in human pancreatic cancer, mainly via the PI3-K/AKT signaling pathway [42]. As a consequence of these findings, it is thought that MMP inhibitors play an important role in human pancreatic cancer. Kilian et al. reported that low-dose therapy with a selective MMP inhibitor (Ro 28-2653) decreases liver metastasis due to an inhibition of MMP-2 and -9 concentrations in ductal pancreatic cancer [43]. BB-94 (batimastat), a bioactive synthetic MMP inhibitor, has been shown to inhibit MMP activity and invasive potential in pancreatic cancer cell lines [44]. Several clinical trials using BB-2516 and BAY 12-9566 have recently been undertaken: however, no significant overall survival advantage was seen in advanced pancreatic cancer patients [45, 46].

Tissue inhibitors of metalloproteinases (TIMPs) are small proteins capable of binding

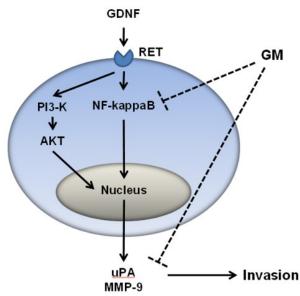


Figure 3. Gabexate mesilate (GM) inhibits the invasive ability of pancreatic cancer cells. GDNF binds the RET tyrosine kinase receptor, and then the NF-kappa B and PI3-K/AKT pathways are activated. We consider gabexate mesilate to be capable of inhibiting pancreatic cancer invasion by antagonizing the activities of uPA and suppressing uPA production.

and inactivating MMPs; four TIMPs have been identified [47, 48]. Boomston *et al.* reported that TIMP-1 overexpression reduces the invasive potential of pancreatic cancer [49] and that TIMP-1 antisense gene transfection cells showed marked reductions in cell invasion and MMP-2 activity [50].

Cysteine Proteases and Their Inhibitors

Cathepsin cysteine proteases have been implicated in processes important for tumor development and progression, including angiogenesis, cell proliferation, apoptosis and invasion [51]. There are 11 cysteine cathepsins present in the human genome and 19 in the mouse genome [52]. It has been noted that invasion is facilitated by a membrane or secreted form of cathepsin B which acts outside the cell to degrade extracellular matrix components at or adjacent to the surface of the invading cell [53]. Cathepsin B is able to degrade the components of the extracellular matrix and basement membrane either directly indirectly by activating other proteases such as pro-uPA [54]. In pancreatic cancer, it is reported that cathepsin B and cathepsin L are

strong and independent prognostic markers [55]. Interruption of their expression, either by antisense RNA [56] or RNA interference [57], was found to reduce tumor cell invasion, angiogenesis and tumor growth. Stefin A is an endogenous inhibitor of cathepsin overexpression of stefin A (as with TIMP) inhibits tumor cell growth, angiogenesis, invasion, and metastasis [58]. The specific cysteine cathepsin B inhibitors in a previous study were purified [59]; however, the biological effects of these drugs on pancreatic cancer are not completely understood. Further investigation about the role of cysteine their inhibitors proteases and prevention of pancreatic cancer invasion and metastasis is needed.

Conclusion

In conclusion, we have demonstrated that GDNF increases NF-kappa B activity in human pancreatic cancer cell lines and that the invasive potential is regulated by NFkappa B activation. Furthermore, the serine protease inhibitor gabexate mesilate may play an important role in the inhibition of neural invasion in human pancreatic cancer cells. Many protease inhibitors strongly influence pancreatic cancer invasion and metastasis. We foresee protease inhibitors eventually becoming part of the paradigm of treatment for pancreatic cancer, thus improving the prognoses of those with resectable and unresectable disease

Keywords Gabexate; Neoplasm Invasiveness; NF-kappa B; Pancreatic Neoplasms; Protease Inhibitors

Abbreviations GDNF: glial cell line-derived neurotrophic factor; HUVEC: umbilical vein endothelial cell: MMP: matrix metalloproteinases; NF-kappa B: nuclear kappaB; PPAR: peroxisome factor proliferators-activated receptor; TAT: tumorassociated trypsinogen; TIMP: tissue inhibitors of metalloproteinases; uPA: urokinase-type plasminogen activator; uPAR: urokinase-type plasminogen activator receptor **Conflict of interest** The authors have no potential conflicts of interest

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References

- 1. Takahashi T, Ishikura H, Motohara T, Okushiba S, Dohke M, Katoh H. Perineural invasion by ductal adenocarcinoma of the pancreas. J Surg Oncol 1997; 65:164-70. [PMID 9236924]
- 2. Hirai I, Kimura W, Ozawa K, Kudo S, Suto K, Kuzu H, Fuse A. Perineural invasion in pancreatic cancer. Pancreas 2002; 24:15-25. [PMID 11741178]
- 3. Ozaki H, Hiraoka T, Mizumoto R, Matsuno S, Matsumoto Y, Nakayama T, et al. The prognostic significance of lymph node metastasis and intrapancreatic perineural invasion in pancreatic cancer after curative resection. Surg Today 1999; 29:16-22. [PMID 9934826]
- 4. Friess H, Yamanaka Y, Büchler M, Ebert M, Beger HG, Gold LI, Korc M. Enhanced expression of transforming growth factor beta isoforms in pancreatic cancer correlates with decreased survival. Gastroenterology 1993; 105:1846-56. [PMID 8253361]
- 5. Lin LF, Doherty DH, Lile JD, Bektesh S, Collins F. GDNF: a glial-cell-line derived neurotrophic factor for midbrain dopaminergic neurons. Science 1993; 260:1130-2. [PMID 8493557]
- 6. Henderson CE, Phillips HS, Pollock RA, Davies AM, Lemeulle C, Armanini M, et al. GDNF: a potent survival factor for motoneurons present in peripheral nerve and muscle. Science 1994; 266:1062-4. [PMID 7973664]
- 7. Okada Y, Takeyama H, Sato M, Morikawa M, Sobue K, Asai K, et al. Experimental implication of celiac ganglionotropic invasion of pancreatic-cancer cells bearing c-ret proto-oncogene with reference to glial-cell-line-derived neurotrophic factor (GDNF). Int J Cancer 1999; 81:67-73. [PMID 10077155]
- 8. Funahashi H, Takeyama H, Sawai H, Furuta A, Sato M, Okada Y, et al. Alteration of Expression by

- Glial Cell Line-Derived Neurotrophic Factor (GDNF) in Human Pancreatic Cancer Cells. Pancreas 2003; 27:190-6. [PMID 12883269]
- 9. Takahashi H, Funahashi H, Sawai H, Sakamoto M, Matsuo Y, Yamamoto M, et al. Glial cell line-derived neurotrophic factor enhances nuclear factor-kappaB activity and invasive potential in human pancreatic cancer cells. Pancreas 2004; 29:22-7. [PMID 15211107]
- 10. Chen HM, Chen JC, Hwang TL, Jan YY, Chen MF. Prospective and randomized study of gabexate mesilate for the treatment of severe acute pancreatitis with organ dysfunction. Hepatogastroenterology 2000; 47:1147-50. [PMID 11020900]
- 11. Cavallini G, Tittobello A, Frulloni L, Masci E, Mariana A, Di Francesco V. Gabexate for the prevention of pancreatic damage related to endoscopic retrograde cholangiopancreatography. Gabexate in digestive endoscopy. Italian Group. N Engl J Med 1996; 335:919-23. [PMID 8786777]
- 12. Aosasa S, Ono S, Mochizuki H, Tsujimoto H, Ueno C, Matsumoto A. Mechanism of the inhibitory effect of protease inhibitor on tumor necrosis factor alpha production of monocytes. Shock 2001; 15:101-5. [PMID 11220636]
- 13. Uchiba M, Okajima K, Kaun C, Binder BR, Wojta J. Gabexate mesilate, a synthetic anticoagulant, inhibits the expression of endothelial leukocyte adhesion molecules in vitro. Crit Care Med 2003; 31:1147-53. [PMID 12682486]
- 14. Wang CY, Mayo MW, Baldwin AS. TNF- and cancer therapy-induced apoptosis: potentiation by inhibition of NF-kappaB. Science 1996; 274:784-7. [PMID 8864119]
- 15. Arlt A, Vorndamm J, Breitenbroich M, Fölsch UR, Kalthoff H, Schmidt WE, Schäfer H. Inhibition of NF-kappaB sensitizes human pancreatic carcinoma cells to apoptosis induced by etoposide (VP16) or doxorubicin. Oncogene 2001; 20:859-68. [PMID 11314019]
- 16. Arlt A, Vorndamm J, Müerköster S, Yu H, Schmidt WE, Fölsch UR, Schäfer H. Autocrine production of interleukin 1beta confers constitutive nuclear factor kappaB activity and chemoresistance in pancreatic carcinoma cell lines. Cancer Res 2002; 62:910-6. [PMID 11830551]
- 17. Huang S, Pettaway CA, Uehara H, Bucana CD, Fidler IJ. Blockade of NF-kappaB activity in human prostate cancer cells is associated with suppression of angiogenesis, invasion, and metastasis. Oncogene 2001; 20:4188-97. [PMID 11464285]
- 18. Huang S, DeGuzman A, Bucana CD, Fidler IJ. Nuclear factor-kappaB activity correlates with growth, angiogenesis, and metastasis of human melanoma cells in nude mice. Clin Cancer Res 2000; 6:2573-81. [PMID 10873114]

- 19. Nakshatri H, Bhat-Nakshatri P, Martin DA, Goulet RJ, Sledge GW. Constitutive activation of NF-kappaB during progression of breast cancer to hormone-independent growth. Mol Cell Biol 1997; 17:3629-39. [PMID 9199297]
- 20. Wang W, Abbruzzese JL, Evans DB, Larry L, Cleary KR, Chiao PJ. The nuclear factor-kappa B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells. Clin Cancer Res 1999; 5:119-27. [PMID 9918209]
- 21. Scaife CL, Kuang J, Wills JC, Trowbridge DB, Gray P, Manning BM, et al. Nuclear factor kappaB inhibitors induce adhesion-dependent colon cancer apoptosis: implications for metastasis. Cancer Res 2002; 62:6870-8. [PMID 12460901]
- 22. Shah SA, Potter MW, McDade TP, Ricciardi R, Perugini RA, Elliott PJ, et al. 26S proteasome inhibition induces apoptosis and limits growth of human pancreatic cancer. J Cell Biochem 2001; 82:110-22. [PMID 11400168]
- 23. Müerköster S, Arlt A, Witt M, Gehrz A, Haye S, March C, et al. Usage of the NF-kappaB inhibitor sulfasalazine as sensitizing agent in combined chemotherapy of pancreatic cancer. Int J Cancer 2003; 104:469-76. [PMID 12584745]
- 24. McDade TP, Perugini RA, Vittimberga FJ, Carrigan RC, Callery MP. Salicylates inhibit NF-kappaB activation and enhance TNF-alpha-induced apoptosis in human pancreatic cancer cells. J Surg Res 1999; 83:56-61. [PMID 10210643]
- 25. DeClerck YA, Imren S. Protease inhibitors: role and potential therapeutic use in human cancer. Eur J Cancer 1994; 30A:2170-80. [PMID 7857719]
- 26. Stetler-Stevenson WG, Aznavoorian S, Liotta LA. Tumor cell interactions with the extracellular matrix during invasion and metastasis. Annu Rev Cell Biol 1993; 9:541-73. [PMID 8280471]
- 27. Wang W, Abbruzzese JL, Evans DB, Chiao PJ. Overexpression of urokinase-type plasminogen activator in pancreatic adenocarcinoma is regulated by constitutively activated RelA. Oncogene 1999; 18:4554-63. [PMID 10467400]
- 28. Cantero D, Friess H, Deflorin J, Zimmermann A, Brundler MA, Riesle E, et al. Enhanced expression of urokinase plasminogen activator and its receptor in pancreatic carcinoma. Br J Cancer 1997; 75:388-95. [PMID 9020484]
- 29. Sawai H, Okada Y, Funahashi H, Matsuo Y, Takahashi H, Takeyama H, Manabe T. Interleukin-lalpha enhances the aggressive behavior of pancreatic cancer cells by regulating the alpha6beta1-integrin and urokinase plasminogen activator receptor expression. BMC Cell Biol 2006; 7:8. [PMID 16504015]

- 30. Sawai H, Liu J, Reber HA, Hines OJ, Eibl G. Activation of peroxisome proliferator-activated receptor-gamma decreases pancreatic cancer cell invasion through modulation of the plasminogen activator system. Mol Cancer Res 2006; 4:159-67. [PMID 16547153]
- 31. Zhang H, Ma G, Dong M, Zhao M, Shen X, Ma Z, Guo K. Epidermal growth factor promotes invasiveness of pancreatic cancer cells through NF-kappaB-mediated proteinase productions. Pancreas 2006; 32:101-9. [PMID 16340751]
- 32. Nawrocki ST, Bruns CJ, Harbison MT, Bold RJ, Gotsch BS, Abbruzzese JL, et al. Effects of the proteasome inhibitor PS-341 on apoptosis and angiogenesis in orthotopic human pancreatic tumor xenografts. Mol Cancer Ther 2002; 1:1243-53. [PMID 12516957]
- 33. Uchima Y, Sawada T, Nishihara T, Maeda K, Ohira M, Hirakawa K. Inhibition and mechanism of action of a protease inhibitor in human pancreatic cancer cells. Pancreas 2004; 29:123-31. [PMID 15257104]
- 34. Takahashi H, Funahashi H, Sawai H, Matsuo Y, Yamamoto M, Okada Y, et al. Synthetic Serine Protease Inhibitor, Gabexate Mesilate, Prevents Nuclear Factor-kappaB Activation and Increases TNF-alpha-Mediated Apoptosis in Human Pancreatic Cancer Cells. Dig Dis Sci 2007; Mar 15. [PMID 17357832]
- 35. Johansson N, Ahonen M, Kähäri VM. Matrix metalloproteinases in tumor invasion. Cell Mol Life Sci 2000; 57:5-15. [PMID 10949577]
- 36. Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. Annu Rev Cell Dev Biol 2001; 17:463-16. [PMID 11687497]
- 37. Cox G, O'Byrne KJ. Matrix metalloproteinases and cancer. Anticancer Res 2001; 21:4207-19. [PMID 11908674]
- 38. Nagase H, Woessner JF. Matrix metalloproteinases. J Biol Chem 1999; 274:21491-4. [PMID 10419448]
- 39. Gress TM, Müller-Pillasch F, Lerch MM, Friess H, Büchler M, Adler G. Expression and in-situ localization of genes coding for extracellular matrix proteins and extracellular matrix degrading proteases in pancreatic cancer. Int J Cancer 1995; 62:407-13. [PMID 7635566]
- 40. Gong YL, Xu GM, Huang WD, Chen LB. Expression of matrix metalloproteinases and the tissue inhibitors of metalloproteinases and their local invasiveness and metastasis in Chinese human pancreatic cancer. J Surg Oncol 2000; 73:95-9. [PMID 10694645]
- 41. Tan X, Egami H, Abe M, Nozawa F, Hirota M, Ogawa M. Involvement of MMP-7 in invasion of

- pancreatic cancer cells through activation of the EGFR mediated MEK-ERK signal transduction pathway. J Clin Pathol 2005; 58:1242-8. [PMID 16311341]
- 42. Okada Y, Eibl G, Duffy JP, Reber HA, Hines OJ. Glial cell-derived neurotrophic factor upregulates the expression and activation of matrix metalloproteinase-9 in human pancreatic cancer. Surgery 2003; 134:293-9. [PMID 12947332]
- 43. Kilian M, Gregor JI, Heukamp I, Hanel M, Ahlgrimm M, Schimke I, et al. metalloproteinase inhibitor RO 28-2653 decreases liver metastasis by reduction of MMP-2 and MMP-9 concentration in BOP-induced ductal pancreatic cancer Syrian Hamsters: inhibition of metalloproteinases in pancreatic cancer. Prostaglandins Leukot Essent Fatty Acids 2006; 75:429-34. [PMID 17034997]
- 44. Zervos EE, Shafii AE, Haq M, Rosemurgy AS. Matrix metalloproteinase inhibition suppresses MMP-2 activity and activation of PANC-1 cells in vitro. J Surg Res 1999; 84:162-7. [PMID 10357914]
- 45. Bramhall SR, Rosemurgy A, Brown PD, Bowry C, Buckels JA; Marimastat Pancreatic Cancer Study Group. Marimastat as first-line therapy for patients with unresectable pancreatic cancer: a randomized trial. J Clin Oncol 2001; 19:3447-55. [PMID 11481349]
- 46. Moore MJ, Hamm J, Dancey J, Eisenberg PD, Dagenais M, Fields A, et al. Comparison of gemcitabine versus the matrix metalloproteinase inhibitor BAY 12-9566 in patients with advanced or metastatic adenocarcinoma of the pancreas: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. J Clin Oncol 2003; 21:3296-302. [PMID 12947065]
- 47. Baker AH, Ahonen M, Kähäri VM. Potential applications of tissue inhibitor of metalloproteinase (TIMP) overexpression for cancer gene therapy. Adv Exp Med Biol 2000; 465:469-83. [PMID 10810650]
- 48. Birkedal-Hansen H, Moore WG, Bodden MK, Windsor LJ, Birkedal-Hansen B, DeCarlo A, Engler JA. Matrix metalloproteinease: a review. Crit Rev Oral Biol Med 1993; 4:197-250. [PMID 8435466]
- 49. Bloomston M, Shafii A, Zervos EE, Rosemurgy AS. TIMP-1 overexpression in pancreatic cancer attenuates tumor growth, decreases implantation and metastasis, and inhibits angiogenesis. J Surg Res 2002; 102:39-44. [PMID 11792150]
- 50. Bloomston M, Shafii A, Zervos E, Rosemurgy AS. TIMP-1 antisense gene transfection attenuates the invasive potential of pancreatic cancer cells in vitro and inhibits tumor growth in vivo. Am J Surg 2005; 189:675-9. [PMID 15910719]
- 51. Gocheva V, Zeng W, Ke D, Klimstra D, Reinheckel T, Peters C, Hanahan D, Joyce JA. Distinct roles for cysteine cathepsin genes in multistage

- tumorigenesis. Genes Dev 2006; 20:543-56. [PMID 16481467]
- 52. Turk V, Kos J, Turk B. Cysteine cathepsins (proteases). On the main stage of cancer? Cancer Cell 2004; 5:409-10. [PMID 15144947]
- 53. Frosch BA, Berquin I, Emmert-Buck MR, Moin K, Sloane BF. Molecular regulation, membrane association and secretion of tumor cathepsin B. APMIS 1999; 107:28-37. [PMID 10190277]
- 54. Kobayashi H, Schmitt M, Goretzki L, Chucholowski N, Calvete J, Kramer M, et al. Cathepsin B efficiently activates the soluble and the tumor cell receptor-bound form of the proenzyme urokinase-type plasminogen activator (Pro-uPA). J Biol Chem 1991; 266:5147-52. [PMID 1900515]
- 55. Niedergethmann M, Wostbrock B, Sturm JW, Willeke F, Post S, Hildenbrand R. Prognostic impact of cysteine proteases cathepsin B and cathepsin L in pancreatic adenocarcinoma. Pancreas 2004; 29:204-11. [PMID 15367886]

- 56. Yanamandra N, Gumidyala KV, Waldron KG, Gujrati M, Olivero WC, Dinh DH, et al. Blockade of cathepsin B expression in human glioblastoma cells is associated with suppression of angiogenesis. Oncogene 2004; 23:2224-30. [PMID 14730346]
- 57. Lakka SS, Gondi CS, Yanamandra N, Olivero WC, Dinh DH, Gujrati M, Rao JS. Inhibition of cathepsin B and MMP-9 gene expression in glioblastoma cell line via RNA interference reduces tumor cell invasion, tumor growth and angiogenesis. Oncogene 2004; 23:4681-9. [PMID 15122332]
- 58. Li W, Ding F, Zhang L, Liu Z, Wu Y, Luo A, et al. Overexpression of stefin A in human esophageal squamous cell carcinoma cells inhibits tumor cell growth, angiogenesis, invasion, and metastasis. Clin Cancer Res 2005; 15:8753-62. [PMID 16361563]
- 59. Greenspan PD, Clark KL, Tommasi RA, Cowen SD, McQuire LW, Farley DL, et al. Identification of dipeptidyl nitriles as potent and selective inhibitors of cathepsin B through structure-based drug design. J Med Chem 2001; 20:4524-34. [PMID 11741472]