

Overexpression of the Sm-Like Proto-Oncogene in Primary and Metastatic Pancreatic Endocrine Tumors

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ABSTRACT

Context The cancer associated Sm-like proto-oncogene mRNA has been found to be overexpressed in the majority of pancreatic adenocarcinomas and is necessary for the transformed phenotype in pancreatic cancer cell lines. However, expression levels have not been examined in other types of pancreatic neoplasms, such as pancreatic endocrine tumors.

Setting Fifteen primary pancreatic endocrine tumors, including five insulinomas and 10 non-functioning tumors, along with seven hepatic metastatic pancreatic endocrine tumors.

Main outcome measures Quantitative expression levels of cancer associated Sm-like mRNA were measured by real-time PCR. Overexpression was defined as a two-fold or greater value when compared to the expression levels found in normal pancreatic islet cells obtained from healthy donors.

Results In primary tumors, four of the 10 non-functioning pancreatic endocrine tumors were found to overexpress cancer associated Sm-like mRNA (40%). Three of the five (60%) insulinomas also overexpressed cancer associated Sm-like mRNA. In total, cancer associated Sm-like mRNA was overexpressed

in seven of 15 primary tumors (47%) and in the majority (71%, 5 of 7) of the hepatic metastases.

Conclusions Our results indicate that the cancer associated Sm-like mRNA gene may also play a role in the tumorigenesis of pancreatic endocrine tumors.

INTRODUCTION

Little is known about the molecular alterations leading to pancreatic endocrine tumor (PET) tumorigenesis. However, the emergence of novel molecular characterization strategies has made it apparent that PETs exhibit diverse molecular fingerprints with respect to tumors arising from the exocrine component of the pancreas [1, 2, 3, 4]. The *p53*, *K-ras*, *p16* and *DPC4* genes are only rarely altered in PETs [2, 5] and, to date, alteration of the *MEN-1* gene remains the most common genetic event noted in gastrinomas and non-functioning PETs [1, 6, 7, 8, 9, 10, 11]. More recently, it has also been shown that non-functioning (NF) PETs may be divided into two molecular phenotypes, distinguishable by ploidy [4]. When utilized in conjunction with the Ki-67 cellular proliferation index, ploidy analysis provides independent statistically significant information which predicts survival [4].

The cancer associated Sm-like (CaSm) gene codes for a protein containing 133 amino acids which contain two Sm motifs found in common small nuclear ribonucleoproteins and has the greatest homology to the SmG protein [12, 13, 14]. CaSm has recently been found to be overexpressed in 87.5% of pancreatic adenocarcinomas when compared to their normal pancreatic tissue and a variety of different tumor types [13]. In addition, the pancreatic adenocarcinoma cell lines, ASPC-1, CAPAN-2, COLO357, PANC-1, CAPAN-1 and HPAC were all found by Northern blot analysis to overexpress CaSm when compared to normal pancreatic tissue [13]. Although the exact mechanism of CaSm proto-oncogene overexpression in oncogenesis is unclear, it is thought that, because of its significant homology to the Sm G protein, it may play a role in RNA splicing [15]. Additional evidence reveals homology of CaSm to the yeast homolog Lsm1 indicating a possible role in mRNA decapping [13, 16]. The importance of CaSm expression in contributing to the transformed state has been manifested by its apparent requirement for transformed pancreatic cell lines to be able to grow in an anchorage-independent manner [13]. In addition, antisense CaSm RNA has been shown to have significant anti-tumor effects at both the cellular and the animal levels [13, 15]. In particular, pancreatic cell lines infected with Ad-alpha-CaSm revealed a statistically significant reduction in proliferation and anchorage independent growth [15]. Using immune-compromized mice with subcutaneous tumor implants, a decrease in tumor growth and increase of median survival was further demonstrated [15]. Because of its potential role in pancreatic oncogenesis, an analysis of Sm-like gene expression levels in PETs is also warranted.

Using real-time PCR, we have examined the expression levels of CaSm in a panel of 15 primary and 7 metastatic PETs, and compared expression levels to those seen in normal islet cells.

MATERIALS AND METHODS

Patients and Tumors

Tumors of 17 patients (8 males and 9 females) were studied. The average age of the patients was 52 years (range 28-75). A total of 15 primary pancreatic tumors (including 10 NF-PET and 5 primary insulinomas) and 7 NF-PET hepatic metastases were studied. Some of these cases have been previously described [4, 17]. Five of the patients with metastatic NF cancers also had the corresponding primary tumor available. One NF tumor and one insulinoma were from patients with MEN-1 syndrome. All tumor samples were collected in the operating room in sterile containers and snap frozen in liquid nitrogen. Histology was confirmed on frozen sections and only tumors of more than 90% tumor cells were used. Normal pancreas and islet cells were from organ donors.

Cell Lines

The pancreatic adenocarcinoma cell lines GER, T3M4, CFPAC1, and PANC-2 were obtained from NR Lemoine (Imperial College School of Medicine, London, UK), PACA3, PACA44 from M von Bülow (University of Mainz, Mainz, Germany), PANC-1, ASPC-1 (American Type Culture Collection, Manassas, VA, USA) and A8184 from H Kalthoff (University Hospital Eppendorf, Hamburg, Germany). All the cell lines were cultured in Roswell Park Memorial Institute (RPMI) 1640 medium supplemented with 10% heat-inactivated fetal calf serum (Gibco-BRL, Life Technologies Inc, Frederick, MA, USA) and were *Mycoplasma* free.

RNA Extraction and cDNA Synthesis

Total RNA from tissue specimens and cell lines was prepared using TRIZOL reagent (Gibco-BRL, Life Technologies Inc, Frederick, MA, USA) according to the manufacturer's protocol. Complementary

DNA was synthesized with the 1st Strand cDNA Synthesis Kit for RT-PCR using avian myeloblastosis virus (AMV) reverse transcriptase and oligo (dT) primers using the manufacturer's recommendations (Roche Molecular Biochemicals, Mannheim, Germany).

Real-Time PCR

The sense and antisense primers used were TACATATGCAACATGAAGAA and CTCTAATTTGGGATATGAAG, respectively, which recognize sequences near the 5' end of the cDNA and amplify a 150-bp fragment. The PCR conditions were 95 °C for 10 min, followed by 40 cycles of 95 °C for 45 sec, 58 °C for 45 sec, and 72 °C for 45 sec. PCR reactions contained 5 µL of SYBER Green PCR Master Mix (PE Biosystems, Foster City, CA, USA), 10 ng of primers and 500 ng of cDNA in a total volume of 10 µL and were performed in triplicate. Beta-actin was also amplified under the same conditions and used to normalize reactions. Sense and antisense primers to the beta-actin gene were TGGAGAAAATCTGGCACCACACC and GATGGGCACAGTGTGGGTGACCC, respectively.

A 9600 thermocycler mounted with an ABI Prism 7700 sequence detection system (Perkin Elmer Biosystems, Foster City, CA, USA), was used to perform the PCR cycles and fluorescence was quantified as a Ct value with Primer Express software (Perkin Elmer Biosystems, Foster City, CA, USA). CaSm expression levels were calibrated using beta-actin expression levels as an internal control. The differences between the mean Ct values of the gene of interest and the housekeeping gene were denoted (delta-Ct) and the difference between delta-Ct and the Ct value of the calibrator sample was labeled delta-delta-Ct. The $\log_2(\text{delta-delta-Ct})$ gave the relative quantitation value of CaSm expression with islet cell expression designated as one. Control wells containing SYBER Green PCR master mix and primers without sample cDNA emitted no fluorescence after 40 cycles.

ETHICS

All the studies performed were approved by the Ethics Committee of Verona University, and the study protocol conforms to the ethical guidelines of the 1975 Declaration of Helsinki.

STATISTICS

Descriptive statistics were evaluated and the frequencies of cases which showed elevated expression levels of CaSm (defined as a two-fold, or greater value when compared to expression levels found in normal pancreas obtained from healthy donors) were reported.

RESULTS

Since it has previously been shown that pancreatic cancer cell lines overexpress CaSm when compared to normal pancreatic tissue [13], a panel of nine pancreatic adenocarcinoma cell-lines was used as positive controls for the ability to detect and quantitate CaSm mRNA levels using real-time PCR. Beta-actin was used to normalize expression levels. The results of this analysis are detailed in Table 1. Five of these cell-lines (56%) showed elevated expression levels (two-fold or greater) when compared to expression levels found in normal pancreas obtained from healthy donors. Thus, these results confirm a previous report which demonstrated elevated expression of CaSm in

Table 1. Relative expression of cancer associated Sm-like proto-oncogene mRNA (CaSm) in pancreatic cancer cell-lines.

Cell-line	Relative CaSm expression ^a
A8184	1.45
ASPC-1	3.00
CFPAC1	0.52
GER	1.73
PACA3	2.99
PACA44	2.69
PANC-1	5.30
PANC-2	5.23
T3M4	1.10

^a Expression relative to that found in normal pancreas obtained from healthy donors

pancreatic cancer cell-lines [13] and further demonstrate the validity of the experimental system.

We next sought to quantify the levels of expression of CaSm in pancreatic endocrine tumors. Pertinent clinicopathological characteristics of the cases under study are detailed in Table 2. A total of 10 NF-PETs, 5 insulinomas and 7 NF hepatic metastases were analyzed. The expression levels of CaSm were normalized to those found in islet cells isolated from a normal pancreas of an organ donor. Four of the 10 NF tumors (40%) and 3 of the 5 (60%) insulinomas showed elevated expression levels of CaSm. The gene was also overexpressed in 5 of the 7 metastatic PETs analyzed (71%).

DISCUSSION

The yearly incidence of pancreatic cancer in the United States is around 30,000 and it is the fifth leading cause of cancer-related death; in Europe, it causes 30,000 deaths per year. The vast majority of pancreatic cancers are ductal in origin with non-ductal pancreatic

cancer comprising about 10% of all tumors diagnosed. Of these, about half are pancreatic endocrine tumors which arise from pancreatic islet cells and may occur either sporadically or as part of multiple endocrine neoplasia type 1 (MEN-1) or von Hippel-Lindau (VHL) syndromes. PETs are traditionally divided into functional and non-functional subtypes depending on whether or not the tumors secrete hormones that give rise to clinical symptoms. Functional PETs are mainly represented by insulinomas, are usually smaller and have a more favorable clinical outcome with respect to NF-PETs; it is possible that this is a consequence of the hormone related symptoms which allow a more timely diagnosis of functional PETs. In fact, the initial clinical presentation of NF-PET is commonly an abdominal mass or any symptom related to the invasion of adjacent structures. PETs are generally slow growing with an average delay in diagnosis of five years and have a higher degree of resectability in contrast to their exocrine counterparts. Because symptoms can usually be controlled pharmacologically (i.e., octreotide,

Table 2. Clinicopathological data and relative expression of cancer associated Sm-like proto-oncogene mRNA (CaSm) in pancreatic endocrine tumors.

Case ^a	Sex	Age	Malignancy	Metastasis	Relative CaSm expression ^d	
					Primary tumor	Metastasis ^e
NF11	Female	52	Malignant	Pancreatic lymph node, liver	NA	2.48
NF13	Female	68	Malignant	Pancreatic lymph node, liver	1.71	NA
NF15 ^b	Female	29	Malignant	Peripancreatic lymph node	2.56	NA
NF16	Male	33	Benign	No	0.84	-
NF37	Male	53	Malignant	Pancreatic lymph node, liver	3.58	3.93
NF45	Female	75	Malignant	Liver	0.98	2.53
NF47	Female	71	Malignant	Liver	0.53	2.96
NF48	Female	39	Malignant	Liver	3.92	11.29
NF49	Female	64	Malignant	Pancreatic lymph node, liver	0.68	1.00
NF50	Female	52	Malignant	Pancreatic lymph node, liver	0.09	NA
NF51	Male	66	Benign	No	10.47	-
NF52	Male	66	Malignant	Pancreatic lymph node, liver	NA	0.19
F14 ^c	Female	62	Benign	No	3.69	-
F15 ^c	Male	56	Benign	No	5.69	-
F16 ^c	Male	28	Benign	No	0.43	-
F17 ^c	Male	33	Benign	No	3.01	-
F18 ^{bc}	Male	31	Benign	No	0.30	-

^a F: functional tumor; NF: non-functioning tumor

^b Multiple endocrine neoplasia type 1 (MEN-1) syndrome

^c Insulinoma

^d Expression relative to that in islet cells obtained from healthy donors

^e Only hepatic metastases analyzed

omeprazole) when tumors are functional, the most common cause of death is from liver failure secondary to hepatic metastasis [18]. As CaSm has been reported to be overexpressed in a wide variety of cancers originating from different tissues including that of the pancreas, liver, ovary, lung and kidney [13], it was of interest to determine if it is also overexpressed in pancreatic tumors other than those arising from the pancreatic ducts. In fact, when grouped together, roughly one-half of all primary PETs revealed overexpression of CaSm. Sixty percent of insulinomas also had CaSm overexpression. Most insulinomas are benign with malignancy rates ranging from 4 to 16% [19] and, in our small panel of five tumors, none were found to have evidence of metastasis or local invasion. Unfortunately, patient NF15 also happened to be one of only two patients with the MEN-1 syndrome making any comments as to the relevance of CaSm expression and this syndrome even more difficult.

Of particular interest, overexpression of the gene was found in 5 of the 7 metastatic PETs. When the tumors from the 5 patients with both primary and metastatic samples were compared, it was noteworthy that all of the secondary tumors expressed more of the proto-oncogene than their primary counterparts (Figure 1). It would be tempting to speculate that overexpression of this gene is more frequent in NF metastatic PETs; however, the number of cases analyzed is too small to make any definitive conclusions. This was exacerbated by the fact that material from 2 patients with hepatic metastases (NF13 and NF50) and one patient with a lymph node metastasis (NF15) was not available for examination. Our results, therefore, seem to indicate that CaSm overexpression may have more of a role in PET tumorigenesis than carcinogenesis. This is reiterated by the observation that patient NF51 has one of the highest levels of CaSm expression yet has no evidence of metastatic disease.

Nonetheless, as CaSm expression appears to be necessary for the maintenance of the transformed state, both in cellular and animal

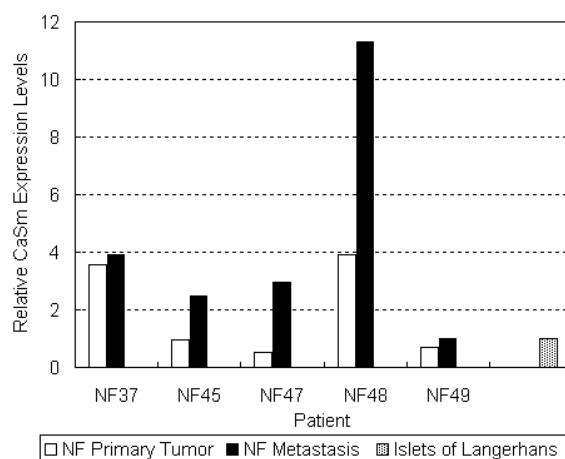


Figure 1. Relative cancer-associated Sm-like proto-oncogene (CaSm) expression from primary and secondary tumors of five patients with non-functioning pancreatic endocrine tumors compared to expression levels from islets of Langerhans from normal donors.

models of pancreatic adenocarcinoma, it has recently been demonstrated that gene therapy approaches using antisense CaSm RNA have antitumor effects [15]. As a result, this gene may represent a novel gene target for new treatment approaches in primary and metastatic PETs that overexpress this proto-oncogene. Studies with a larger patient population are needed to better assess this; however, the rarity of these tumors currently limits our ability to do so. Future studies in CaSm protein production in PETs and other pancreatic tumors may help further clarify the role of this gene in pancreatic oncogenesis.

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Abbreviations AMV: avian myeloblastosis virus; CaSm: cancer associated Sm-like; MEN-1: multiple endocrine neoplasia type 1; NF: non-functioning; PET: pancreatic endocrine tumor; RPMI: Roswell Park Memorial Institute; VHL: von Hippel-Lindau.

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References

1. Moore PS, Missiaglia E, Antonello D, Zamo A, Zamboni G, Corleto V, et al. Role of disease-causing genes in sporadic pancreatic endocrine tumors: MEN1 and VHL. *Genes Chromosomes Cancer* 2001; 32:177-81. [AN 21433812]
2. Moore PS, Orlandini S, Zamboni G, Capelli P, Rigaud G, Falconi M, et al. Pancreatic tumours: molecular pathways implicated in ductal cancer are involved in ampullary but not in exocrine nonductal or endocrine tumorigenesis. *Br J Cancer*. 2001; 84:253-62. [AN 21094882]
3. Moore PS, Zamboni G, Brighenti A, Lissandrini D, Antonello D, Capelli P, et al. Molecular characterization of pancreatic serous microcystic adenomas: evidence for a tumor suppressor gene on chromosome 10q. *Am J Pathol* 2001; 158:317-21. [AN 20580919]
4. Rigaud G, Missiaglia E, Moore PS, Zamboni G, Falconi M, Talamini G, et al. High resolution allelotyping of nonfunctional pancreatic endocrine tumors: identification of two molecular subgroups with clinical implications. *Cancer Res* 2001; 61:285-92. [AN 21036722]
5. Pellegata NS, Sessa F, Renault B, Bonato M, Leone BE, Solcia E, Ranzani GN. K-ras and p53 gene mutations in pancreatic cancer: ductal and nonductal tumors progress through different genetic lesions. *Cancer Res* 1994; 54:1556-60. [AN 94184977]
6. Cupisti K, Hoppner W, Dotzenrath C, Simon D, Berndt I, Roher HD, Goretzki PE. Lack of MEN1 gene mutations in 27 sporadic insulinomas. *Eur J Clin Invest* 2000; 30:325-9. [AN 20223291]
7. Gortz B, Roth J, Krahenmann A, de Krijger RR, Muletta-Feurer S, Rutimann K, et al. Mutations and allelic deletions of the MEN1 gene are associated with a subset of sporadic endocrine pancreatic and neuroendocrine tumors and not restricted to foregut neoplasms. *Am J Pathol* 1999; 154:429-36. [AN 99149647]
8. Hessman O, Lindberg D, Skogseid B, Carling T, Hellman P, Rastad J, et al. Mutation of the multiple endocrine neoplasia type 1 gene in nonfamilial, malignant tumors of the endocrine pancreas. *Cancer Res* 1998; 58:377-9. [AN 98117278]
9. Shan L, Nakamura Y, Nakamura M, Yokoi T, Tsujimoto M, Arima R, et al. Somatic mutations of multiple endocrine neoplasia type 1 gene in the sporadic endocrine tumors. *Lab Invest* 1998; 78:471-5. [AN 98224562]
10. Wang EH, Ebrahimi SA, Wu AY, Kashefi C, Passaro E Jr, Sawicki MP. Mutation of the MENIN gene in sporadic pancreatic endocrine tumors. *Cancer Res* 1998; 58:4417-20. [AN 98438062]
11. Zhuang Z, Vortmeyer AO, Pack S, Huang S, Pham TA, Wang C, et al. Somatic mutations of the MEN1 tumor suppressor gene in sporadic gastrinomas and insulinomas. *Cancer Res* 1997; 57:4682-6. [AN 98014544]
12. Hermann H, Fabrizio P, Raker VA, Foulaki K, Hornig H, Brahms H, Luhrmann R. snRNP Sm proteins share two evolutionarily conserved sequence motifs which are involved in Sm protein-protein interactions. *EMBO J* 1995; 14:2076-88. [AN 95262647]
13. Schweinfest CW, Graber MW, Chapman JM, Pappas TS, Baron PL, Watson DK. CaSm: an Sm-like protein that contributes to the transformed state in cancer cells. *Cancer Res* 1997; 57:2961-5. [AN 97373726]
14. Seraphin B. Sm and Sm-like proteins belong to a large family: identification of proteins of the U6 as well as the U1, U2, U4 and U5 snRNPs. *EMBO J* 1995; 14:2089-98. [AN 95262648]
15. Kelley JR, Brown JM, Frasier MM, Baron PL, Schweinfest CW, Vournakis JN, et al. The cancer-associated Sm-like oncogene: a novel target for the gene therapy of pancreatic cancer. *Surgery*. 2000; 128:353-60. [AN 20382854]
16. Boeck R, Lapeyre B, Brown CE, Sachs AB. Capped mRNA degradation intermediates accumulate

in the yeast *spb8-2* mutant. *Mol Cell Biol* 1998; 18:5062-72. [AN 98378518]

17. Beghelli S, Pelosi G, Zamboni G, Falconi M, Iacono C, Bordi C, Scarpa A. Pancreatic endocrine tumours: evidence for a tumour suppressor pathogenesis and for a tumour suppressor gene on chromosome 17p. *J Pathol* 1998; 186:41-50. [AN 99092298]

18. Lamberts SW, van der Lely AJ, de Herder WW, Hofland LJ. Octreotide. *N Engl J Med* 1996; 334:246-54. [AN 96132562]

19. Solcia E, Sessa F, Rindi G, Villani L, Riva C, Buffa R, Capella C. Classification and histogenesis of gastroenteropancreatic endocrine tumours. *Eur J Clin Invest* 1990; 20 Suppl 1: S72-81. [AN 91092372]